

Structural Data: Introduction to X-ray Crystallography & Cryo-EM

Jesse Rinehart, PhD CBB 752, Spring 2017





Thank you to Yong Xiong and Fred Sigworth for contributions to this lecture

Yale Structure Courses:

C&MP 710b/MB&B 710b4, Electron Cryo-Microscopy for Protein Structure Determination Fred J. Sigworth, C.V. Sindelar

MB&B 720a, Macromolecular Structure and Biophysical Analysis Yong Xiong, Andrew Miranker, Anna Marie Pyle

MB&B 721b, Macromolecular Interactions and Dynamic Properties Anna Pyle, Donald Engelman, Elizabeth Rhoades, Hongwei Wang

MB&B 760b3: Principles of Macromolecular Crystallography
Thomas Steitz

MB&B 761b4: X-ray Crystallography Workshop Yong Xiong, Yorgo Modis, and staff

Pharmacology 529b: Structural Pharmacology Ya Ha, Titus Boggon

Additional Resources:

"Crystallography Made Crystal Clear: A Guide for Users of Macromolecular Models" by Gale Rhodes (Third Edition, 2006 Elsevier/Academic Press)

"Crystallography 101" http://www.ruppweb.org/Xray/101index.html

"Single particle electron cryomicroscopy: trends, issues and future perspective."

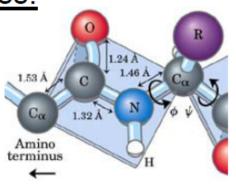
Vinothkumar KR, Henderson R. Q Rev Biophys. 2016 pubmed:27658821

"Just as we see objects around us by interpreting the light reflected from them, x-ray crystallographers "see" molecules by interpreting x-rays diffracted from them." - Gale Rhodes

- There's a <u>limit</u> to how small an object can be seen under a light microscope.
- The diffraction limit: you can not image things that are much smaller than the wavelength of the light you are using.
- The wavelength for visible light is measured in hundreds of nanometers, while atoms are separated by distances of the order of 0.1nm, or 1Å.

We need to use x-rays to resolve atomic features.

Distances between atoms are small: Lab x-ray sources use $CuK\alpha$ radiation. Wavelength = 1.54 Å. Synchrotron radiation wavelengths in the range 0.5 Å - 2.5 Å.

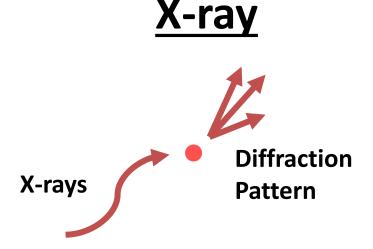


The 2014 Nobel Prize in Chemistry: Eric Betzig, W.E. Moerner, and Stefan Hell "The development of super-resolved fluorescence microscopy"

Spatial Resolution of Biological Imaging Techniques Virus Protein Bacteria Cell Hair Small Ant Mouse Mouse Molecule Brain 10 nm 100 nm 1 µm 10 µm 100 µm 10 cm 1 nm 1 mm 1 cm MRI and Ultrasound Fluorescence Optical Coherence Tomography Microscopy 1Å = 0.1nmWidefield and TIRF Microscopy Superresolution Confocal Microscopy 4Pi and I5M High Resolution Structured Illumination Ground State Depletion (GSD) Saturated Structured Illumination (SSIM) Stimulated Emission Depletion (STED PALM, FPALM and STORM Near-Field (NSOM) Electron Microscopy

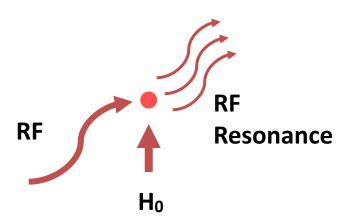
Figure 1

Experimental Determination of Atomic Resolution Structures



- Direct detection of atom positions
- **≻Crystals**

NMR

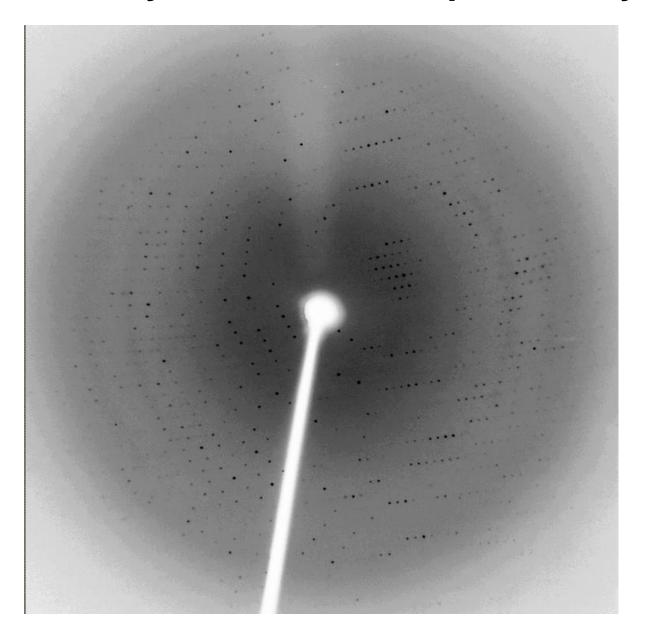


- ➤Indirect detection of H-H distances
- >In solution

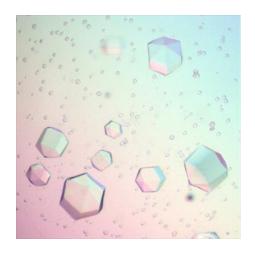
Other methods for determining protein structures:

-EM (Electron Microscopy), Cryo-EM, ESR/Fluorescence

Image of X-ray diffraction of a protein crystal

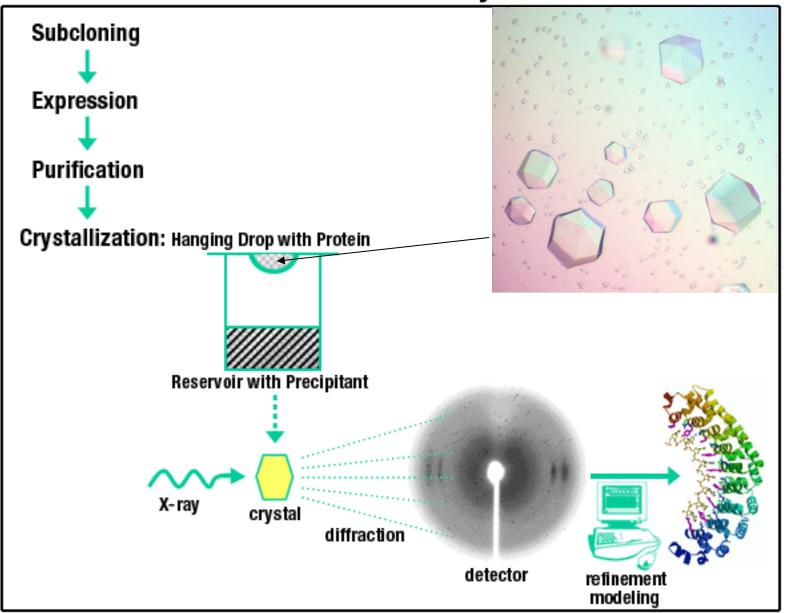


Why Crystals?

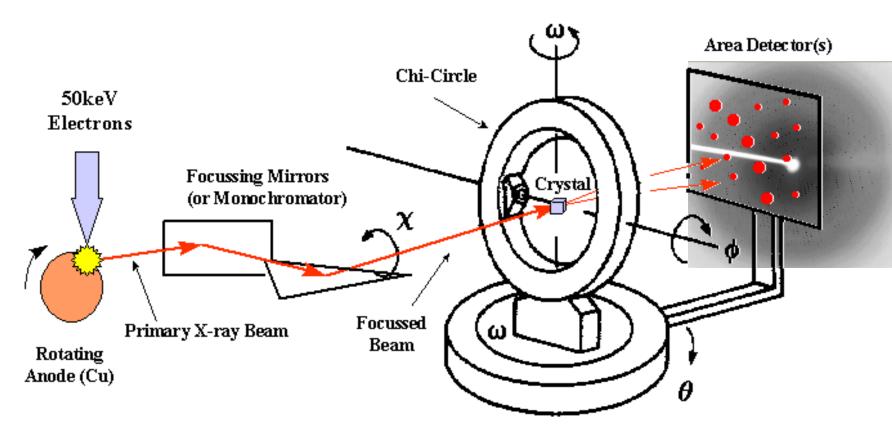


X-rays are scattered by electrons, too weak to record scattering from a single molecule. Crystals are therefore used because they present many molecules (N) in exactly the same orientation. The scattering from each of the N molecules interferes constructively to give a measurable diffraction pattern (enhanced \sim N² fold).

Determination of Protein Crystal Structure



Data Collection



4-Circle Gonoimeter (Eulerian or Kappa Geometry)

Synchrotron X-ray Sources are the method of choice

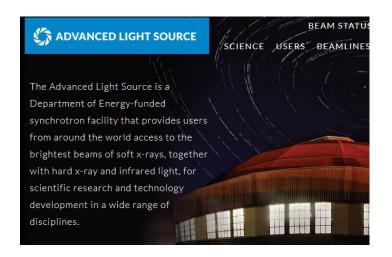
Lab x-ray sources @ 1.54 Å compared to Synchrotron X-ray @ 0.5 Å - 2.5 Å.



APS Chicago



NSLS-II Brookhaven

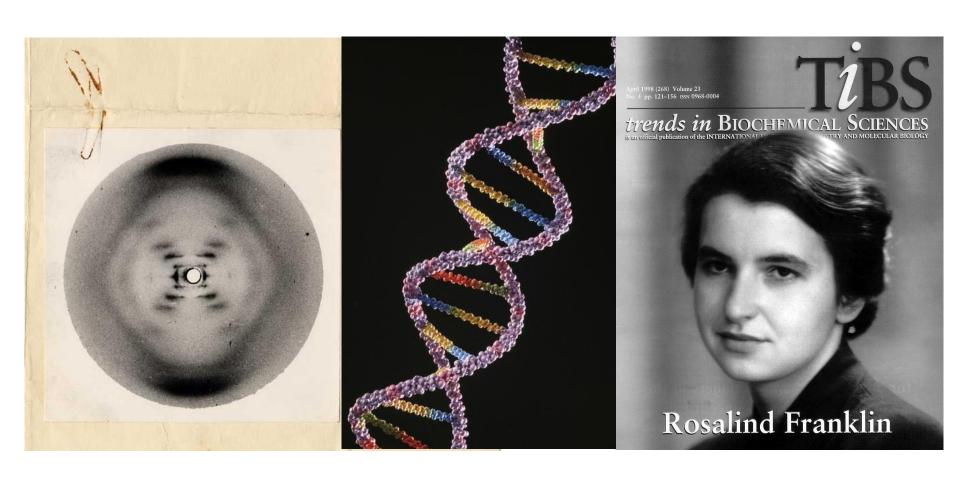


ALS Berkeley

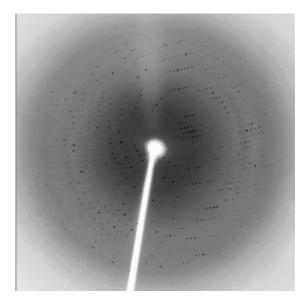


CHESS Ithaca

Most famous X-ray diffraction pattern

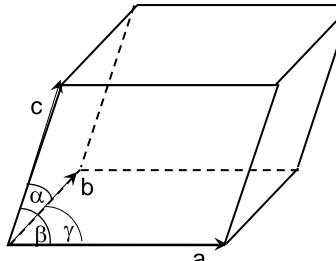


The information we get from a single diffraction experiment



Analyze the pattern of the reflections

- (a) space group of the crystal
- (b) unit cell dimensions



Cubic

$$a = b = c$$
,
 $\alpha = \beta = \gamma = 90^{\circ}$

Hexagonal

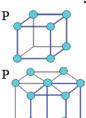
 $\alpha = \beta = 90^{\circ}, \gamma = 120^{\circ}$

Trigonal

$$a = b \neq c$$
,
 $\alpha = \beta = 90^{\circ}$, $\gamma = 120^{\circ}$

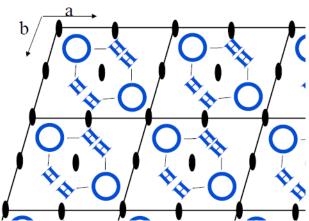
Tetragonal

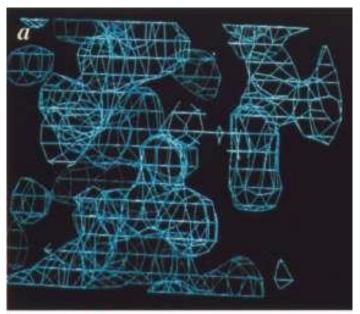




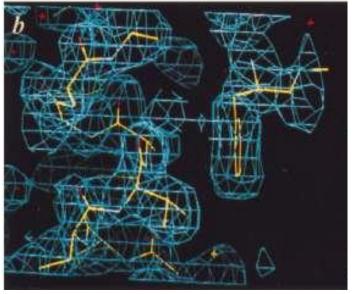
How to understand symmetry? Crystal = lattice + unit cell content

(asymmetric units (asu) content)





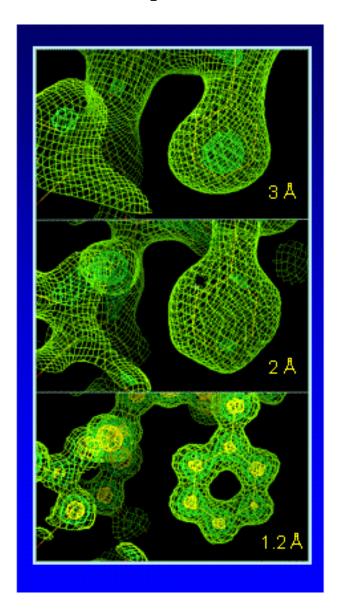
Electron density map

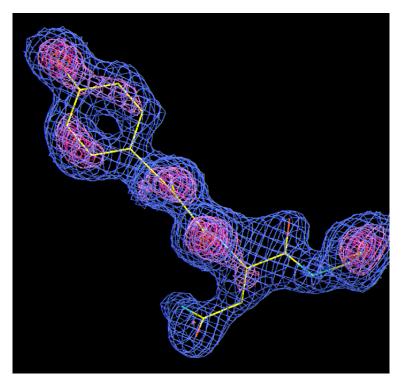


Building a structure model

- © 2006
- Academic Press

The importance of resolution





Experimental electron density map created from multi-wavelength data collected at SSRL beam line 1-5 on a Gold derivative of tetanus C fragment.

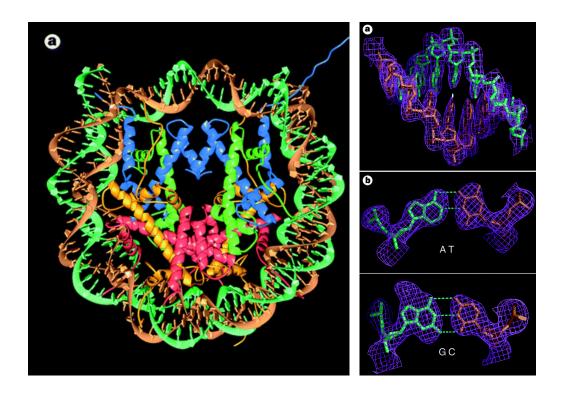
Example of high quality Experimental data where very little refinement has been applied to fit a tyrosine into the density map.

Crystal structure of the nucleosome core particle at 2.8 Å resolution

Karolin Luger, Armin W. Mäder, Robin K. Richmond, David F. Sargent & Timothy J. Richmond

Institut für Molekularbiologie und Biophysik ETHZ, ETH-Hönggerberg, CH-8093 Zürich, Switzerland

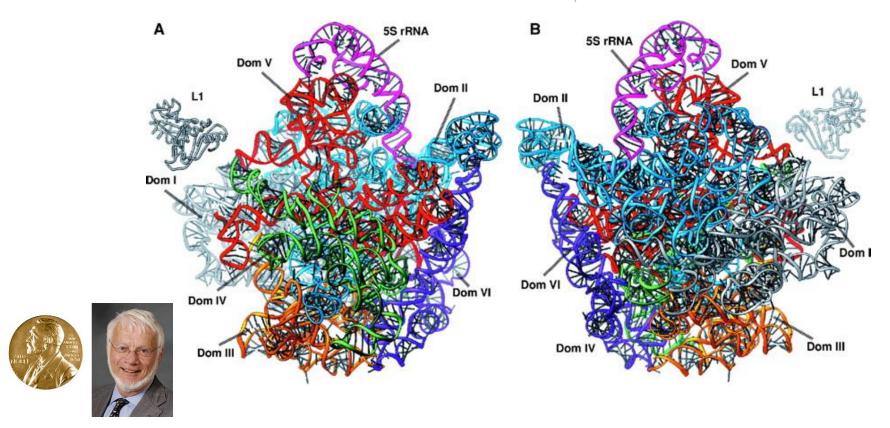
The X-ray crystal structure of the nucleosome core particle of chromatin shows in atomic detail how the histone protein octamer is assembled and how 146 base pairs of DNA are organized into a superhelix around it. Both histone/histone and histone/DNA interactions depend on the histone fold domains and additional, well ordered structure elements extending from this motif. Histone amino-terminal tails pass over and between the gyres of the DNA superhelix to contact neighbouring particles. The lack of uniformity between multiple histone/DNA-binding sites causes the DNA to deviate from ideal superhelix geometry.





The Complete Atomic Structure of the Large Ribosomal Subunit at 2.4 Å Resolution

Nenad Ban, 1* Poul Nissen, 1* Jeffrey Hansen, 1 Peter B. Moore, 1,2
Thomas A. Steitz 1,2,3 †



Thomas Steitz shared 2009 Nobel Prize in Chemistry for this structure

Protein Structure Databases

- Where does protein structural information reside?
 - PDB:
 - http://www.rcsb.org/pdb/
 - MMDB:
 - http://www.ncbi.nlm.nih.gov/Structure/
 - FSSP:
 - http://www.ebi.ac.uk/dali/fssp/
 - SCOP:
 - http://scop.mrc-lmb.cam.ac.uk/scop/
 - CATH:
 - http://www.biochem.ucl.ac.uk/bsm/cath_new/

of structures

2014: 97,180

2016: 115,559

2017: 117,184

http://www.rcsb.org/pdb/home/home.do

RCSB PDB

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An Information Portal to 115559 Biological

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Macromolecular Structures











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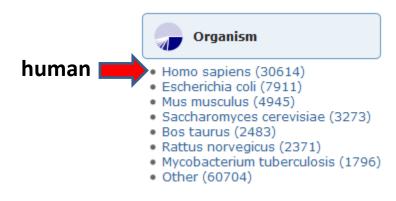


A Structural View of Biology

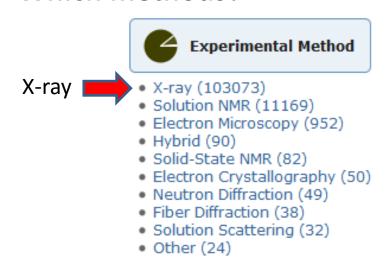
This resource is powered by the Protein Data Bank archive-information about the 3D shapes of proteins, nucleic acids, and complex assemblies that helps students and researchers understand all aspects of biomedicine and agriculture, from protein synthesis to health and disease

January Molecule of the Month

PDB: What species are the structures from?

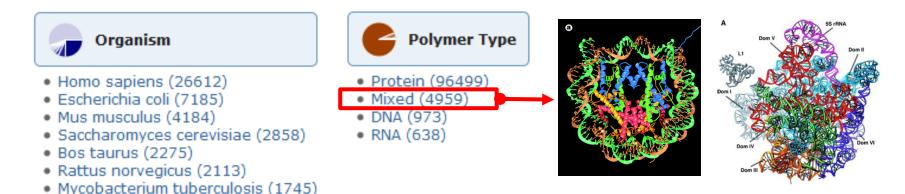


Which methods?



PDB X-ray Structures:

http://www.rcsb.org/pdb/results/results.do?outformat=&grid=1B04C26E&tabtoshow=Current





Membrane Proteins

Small % of the total x-ray data

- ALPHA-HELICAL (2100)
- MONOTOPIC MEMBRANE PROTEINS (358)
- BETA-BARREL (352)

Other (55853)

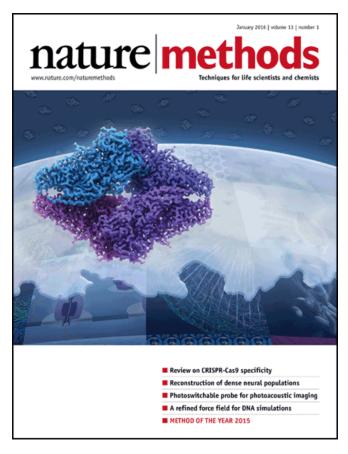
- Jmol
 - http://jmol.sourceforge.net
- **PyMOL**
 - http://pymol.sourceforge.net
- **Swiss PDB viewer**
 - http://www.expasy.ch/spdbv

Tools for Viewing Structures

Mage/KiNG

- http://kinemage.biochem.duke.edu/software/mage.php
- http://kinemage.biochem.duke.edu/software/king.php
- Rasmol
 - http://www.umass.edu/microbio/rasmol/

2015 Method of the Year: Single-particle Cryo-EM



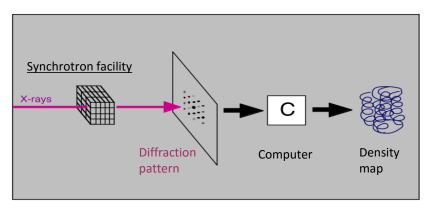
NATURE METHODS | VOL.13 NO.1 | JANUARY 2016

METHOD OF THE YEAR 2015

At *Nature Methods* we are ringing in a new year with our celebration of single-particle cryoelectron microscopy (cryo-EM) as our Method of the Year 2015. Cryo-EM has its roots in work first performed in the 1960s. It has steadily progressed over the past few decades as a medium-resolution structural technique for obtaining information about macromolecular samples that resist analysis by X-ray crystallography. But very recent technical advances, especially the development of direct-detection cameras, have enabled the field to achieve impressive leaps in resolution even reaching the near-atomic realm of X-ray crystallography—and, by extension, biological applicability. An Editorial, News Feature, Primer, Historical Commentary and Commentary discuss how cryo-EM works, what it is used for, how the field began, why now is such an exhilarating time, and where the field is going in the future. We also cast our predictions about methods with exciting potential in our Methods to Watch section.

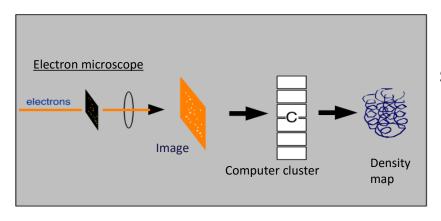
Special feature starts on p19

Two methods for structure determination



X-ray crystallography

Well-established (since 1960s) Requires well-ordered crystals >10¹² copies of protein



Single-particle cryo-EM

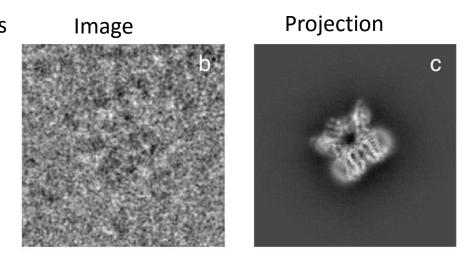
Recent (1990s-present) No crystals required! ~10⁵ copies of protein

The Cryo-EM specimen gives only a phase contrast image

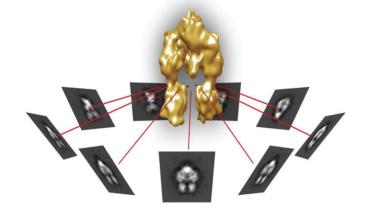
A constellation of images and data processing are essential.

1/4 of a micrograph, showing some particles

Y. Cheng and D. Julius lab. Nature 2013



- orientation assignment and averaging
- 3D reconstruction

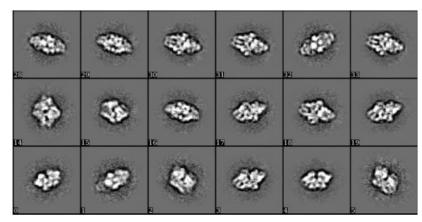


A landmark study for high-resolution single-particle structures

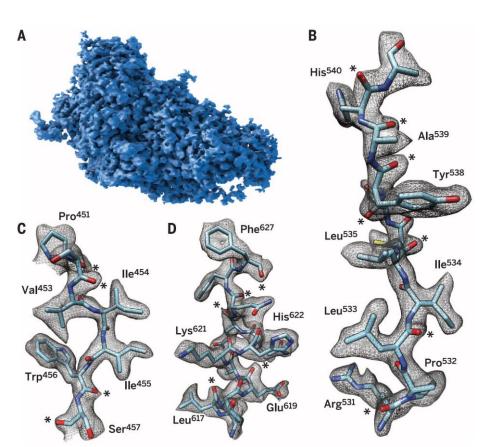
2.2 Å resolution cryo-EM structure of β -galactosidase in complex with a cell-permeant inhibitor

Alberto Bartesaghi, 1* Alan Merk, 1* Soojay Banerjee, 1 Doreen Matthies, 1 Xiongwu Wu, 2 Jacqueline L. S. Milne, 1 Sriram Subramaniam 1 †

Science 2015



2D class averages



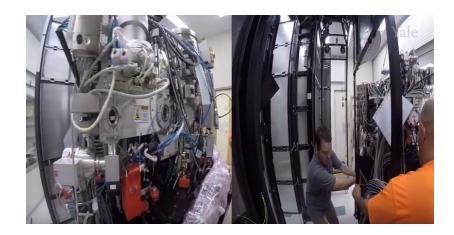
New Technologies, Automation, & Computation are accelerating the field



Krios at National University of Singapore

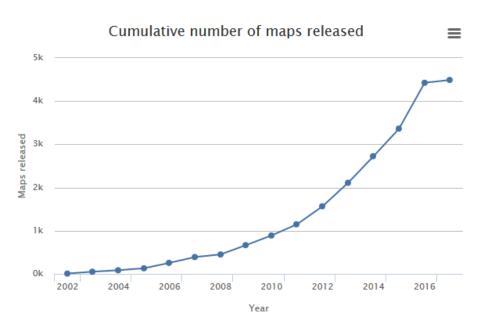


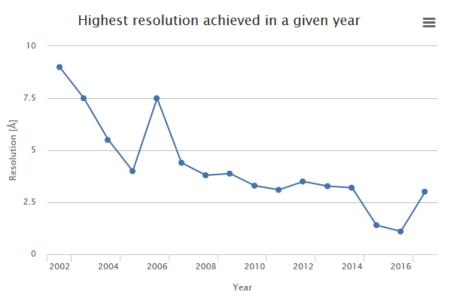
Control room at Scripps Research Institute, La Jolla



Krios TEM installation on Yale's West Campus. Online Summer 2017

EMDB statistics





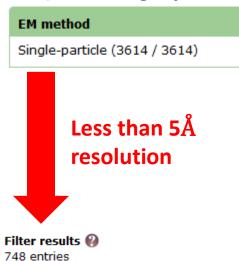




EM resources

https://www.ebi.ac.uk/pdbe/emdb/

Jan 2017: 4,492 total maps 3,614 (~77% single particle)



Component type

Protein (2603 / 2603)

Virus (761 / 761)

Nucleic acid (415 / 415)

Ligand (274 / 274)

Prokaryotic ribosome (264 / 264)

Eukaryotic ribosome (217 / 217)

Cell component (113 / 113)

EM label (2 / 2)

EM method

Single-particle (661 / 661)

Helical (63 / 63)

2D crystallography (21 / 21)

Subtomogram averaging (3 / 3)

Component type

Protein (601 / 601)

Ligand (178 / 178)

Nucleic acid (140 / 140)

Virus (134 / 134)

Eukaryotic ribosome (69 / 69)

Cell component (44 / 44)

Prokaryotic ribosome (22 / 22)